

Pharmacology & Therapeutics 95 (2002) 203-220



Cannabinoids in the treatment of glaucoma

Tomi Järvinen^{a,*}, David W. Pate^b, Krista Laine^a

^aDepartment of Pharmaceutical Chemistry, University of Kuopio, P.O. Box 1627, FIN-70211 Kuopio, Finland ^bHortaPharm B.V., Amsterdam, The Netherlands

Abstract

The leading cause of irreversible blindness is glaucoma, a disease normally characterized by the development of ocular hypertension and consequent damage to the optic nerve at its point of retinal attachment. This results in a narrowing of the visual field, and eventually results in blindness. A number of drugs are available to lower intraocular pressure (IOP), but, occasionally, they are ineffective or have intolerable side-effects for some patients and can lose efficacy with chronic administration. The smoking of marijuana has decreased IOP in glaucoma patients. Cannabinoid drugs, therefore, are thought to have significant potential for pharmaceutical development. However, as the mechanism surrounding their effect on IOP initially was thought to involve the CNS, issues of psychoactivity hindered progress. The discovery of ocular cannabinoid receptors implied an explanation for the induction of hypotension by topical cannabinoid applications, and has stimulated a new phase of ophthalmic cannabinoid research. Featured within these investigations is the possibility that at least some cannabinoids may ameliorate optic neuronal damage through suppression of *N*-methyl-D-aspartate receptor hyperexcitability, stimulation of neural microcirculation, and the suppression of both apoptosis and damaging free radical reactions, among other mechanisms. Separation of therapeutic actions from side-effects now seems possible through a diverse array of novel chemical, pharmacological, and formulation strategies.

© 2002 Elsevier Science Inc. All rights reserved.

Keywords: Cannabinoids; Endocannabinoids; CB receptor; Intraocular pressure; Glaucoma; Topical

Abbreviations: AEA, arachidonylethanolamide; 2-AG, 2-arachidonoylglycerol; CD, cyclodextrin; COX, cyclo-oxygenase; FAAH, fatty acid amide hydrolase; HP-β-CD, hydroxypropyl-β-cyclodextrin; IOP, intraocular pressure; NMDA, *N*-methyl-D-aspartate; P_{app}, apparent partition coefficient; PMSF, phenyl-methylsulfonyl fluoride; SBE-β-CD, sulfobutylether-β-cyclodextrin; THC, tetrahydrocannabinol; VR1, vanilloid receptor Type 1.

Contents

1.	Introd	uction	204
2.	Ocula	r drug delivery	204
	2.1.	Pre-corneal drug elimination	205
	2.2.		205
	2.3.	Physicochemical properties	205
3.			206
	3.1.		206
	3.2.	Ophthalmic applications	207
	3.3.	Cannabinoid administration	208
4.	Ocula	r cannabinoid systems	208
	4.1.	Endogenous cannabinoids	208
	4.2.		209
5.		binoids and intraocular pressure	209

0163-7258/02/\$ – see front matter © 2002 Elsevier Science Inc. All rights reserved. PII: \$0163-7258(02)00259-0

•

^{*} Corresponding author. Tel.: +358-17-162468; fax: +358-17-162456. *E-mail address*: tomi.jarvinen@uku.fi (T. Järvinen).

	5.1.	Classica	al cann	abinoids	s				 			 						 	210
		5.1.1.	Syste	mic adn	ninistı	ratio	1		 			 						 	210
		5.1.2.		nalmic a															210
	5.2.	Nonclas																	211
	5.3.			doles															211
	5.4.	Endoca																	212
		5.4.1.		nidonyle															212
		5.4.2.		ostituted															212
		5.4.3.	Anan	damide	uptak	e inl	nibi	tors	 			 						 	212
	5.5.	Bimato																	213
	5.6.	HU-211																	213
6.	Mecha	anisms of																	213
	6.1.			essure re															213
	6.2.	Neurolo																	214
7.	Canna	binoids																	215
	7.1.	Optima																	215
	7.2.	CB_2 rec																	215
	7.3.	Cannab																	215
8.		usions .																	215
		gements																	216
																			216
1/010	1011003								 	 •	 •	 	•	 •			•	 	210

1. Introduction

Glaucoma is the leading cause of irreversible blindness, and it is estimated that 66.8 million people are presently affected, of whom, 6.7 million will become blind in both eyes (Alward, 1998). Current glaucoma treatment includes α_2 -adrenoreceptor agonists, β -adrenoreceptor agonists, dopaminergic agonists, cholinergic agonists, carbonic anhydrase inhibitors, and prostaglandin agonists, all of which are ocular hypotensive agents (Sugrue, 1997).

Hepler and Frank (1971) originally observed that subjects who smoked marijuana developed a reduced intraocular pressure (IOP). Subsequent human experiments involving oral (Merritt et al., 1980b) and intravenous (Cooler & Gregg, 1977) administration of pure Δ^9 -tetrahydrocannabinol (THC), the main psychoactive ingredient in *Cannabis sativa* L., confirmed this observation and pinpointed the active component. As a result, a great deal of research exploring this compound, and related compounds, as a possible glaucoma drug(s) has been published over the last 20 years (e.g., Green, 1998, 2000; Green & McDonald, 1987). However, clinical application of Δ^9 -THC may include adverse psychic or somatic side-effects, in addition to the fact that the lipophilic cannabinoids are insoluble in water, hindering their use as topical agents.

Discovery of a cannabinoid receptor (Devane et al., 1988) and its endogenous ligand (Devane et al., 1992b), arachidonylethanolamide (AEA), offered new horizons for the use of cannabinoids in medicine, including glaucoma therapy. The presence of CB₁ receptors (Porcella et al., 1998; Straiker et al., 1999a, 1999b) and an AEA-specific enzyme activity in the eye (Matsuda et al., 1997) provided

the context for a mechanism of drug action. In addition, novel formulation technologies, such as cyclodextrins (CDs) (Jarho et al., 1998) or microemulsions (Muchtar et al., 1992), enable the preparation of topical dosage forms for these highly lipophilic cannabinoids. Topically administered cannabinoids having optimal ophthalmic delivery properties would minimize drug concentrations in systemic circulation and possible consequent adverse side-effects.

This review will focus on the cannabinoids as a potential class of topical anti-glaucoma agents, apparently acting upon a newly discovered ocular cannabinoid receptor to lower the ocular hypertension symptomatic of the disease. In addition, mention will be made of possible complimentary mechanisms by which this class of drugs may also offer unique advantages for ameliorating the collateral neuro-degenerative effects of this disease. In addition, a brief summary concerning the main features of drug delivery after ocular administration and CD technology in ophthalmic applications is described in order to clarify future challenges in the research and development of novel ophthalmic cannabinoids.

2. Ocular drug delivery

Topical delivery of eye drops into the lower cul-de-sac of the eye is the most common method of drug treatment in ocular disease. In general, the site of action for ophthalmic drugs is located inside the eye. Unfortunately, after instillation of an eyedrop, typically < 5% of an applied dose reaches the intraocular tissues. The main reason of this low ocular drug availability is poor drug penetration across the

corneal barrier and a rapid loss of the instilled solution from the pre-corneal area. Ocular absorption and pharmacokinetics have been described in numerous reviews (e.g., Davies, 2000; Järvinen, K. et al., 1995; Sasaki et al., 1999). Only the most salient features of ophthalmic drug delivery are summarized herein to help readers understand the process of drug development, especially as applied to cannabinoids.

2.1. Pre-corneal drug elimination

After topical administration, aqueous eyedrop solutions mix with tear fluid and are dispersed over the eye surface. However, various pre-corneal factors (i.e., drainage of instilled solution, non-corneal absorption, induced lacrimation) limit ocular absorption by shortening the cornea contact time of applied drugs. These factors, and the corneal barrier itself, limit penetration of a topically administered ophthalmic drug. As a result, only a few percent of the applied dose is delivered into the intraocular tissues, the major part (50–99%) being absorbed into the systemic circulation (Fig. 1), where it can cause various side-effects.

The main sites for systemic absorption are the nasal mucosa and the ocular conjunctiva (Urtti & Salminen, 1993). Following instillation of an applied eyedrop (25–50 μ L) onto the pre-corneal area of the eye, the greater part of the drug solution rapidly exits the eye surface via the lacrimal drainage system, nasal mucosa, and pharynx. Soon thereafter, the resident tear volume of 7.5 μ L returns to normal (Chrai et al., 1973). Compared with the cornea, the conjunctiva is highly vascularized, with a 10-fold greater surface area (Watsky et al., 1988). It is also 2–30 times

more permeable, depending on the administered drug (Ahmed et al., 1987; Wang et al., 1991).

2.2. Corneal barriers

The cornea is generally considered to be a major, but not exclusive, pathway for the ocular penetration of topically applied drugs (Doane et al., 1978). Compared with many other epithelial tissues (e.g., bronchial, intestinal, nasal, tracheal), the corneal epithelium is relatively impermeable, but less so than the stratum corneum of the skin (Rojanasakul et al., 1992). Although the cornea is composed of five layers, the epithelium and the stroma are most significant for drug delivery (Huang et al., 1983). The lipophilic epithelium is the primary barrier for corneal permeation by highly hydrophilic drugs. However, for highly lipophilic drugs, partitioning from the epithelium to the hydrophilic stroma is the rate-limiting step.

The apparent corneal permeability coefficient (or flux) is usually determined using an isolated cornea mounted in a side-by-side diffusion cell. In vitro corneal permeability studies produce information about the effects of drug structure and formulation on corneal permeability. However, in vitro corneal permeability studies do not include precorneal loss processes and, therefore, do not reliably predict the in vivo bioavailability of topically administered drugs.

2.3. Physicochemical properties

Lipophilic drugs penetrate the corneal epithelium via the transcellular pathway and hydrophilic molecules utilize the paracellular route, the latter involving passive or altered

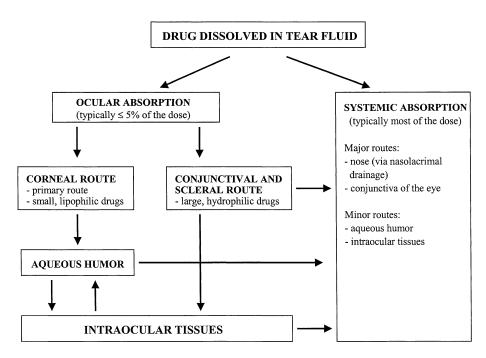


Fig. 1. Main absorption routes of a topically applied ophthalmic drug.

diffusion through intercellular spaces (Borchardt, 1990). For most topically applied drugs, this passive diffusion along a concentration gradient, which is largely influenced by various physicochemical properties, is the main corneal permeation mechanism.

Drug lipophilicity seems to be the most important property for corneal penetration, and both parabolic (Chien et al., 1991) and sigmoidal (Wang et al., 1991) curves have been used to describe their relationship. The optimum apparent partition coefficient (P_{app}; octanol/pH 7.4 buffer) for corneal drug absorption is in the range of 100–1000 (Schoenwald & Huang, 1983; Schoenwald & Ward, 1978), which is consistent with the lipophilic nature of the corneal epithelium (Sasaki et al., 1999).

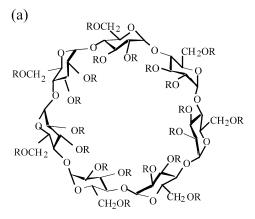
Aqueous solubility is another drug property important for efficacy of delivery. The surface of the eye is constantly being cleaned and moistened by the aqueous tear fluid. Thus, it is difficult for drug molecules to be absorbed by the corneal epithelium, unless they are soluble in the tear film (Loftsson & Stefansson, 1997). In addition, the water solubility of the drug must be good enough to enable the formulation of aqueous eyedrops. The dilemma is that an ideal potential ophthalmic drug should simultaneously be both water-soluble and lipid soluble, but only a few molecules can fulfill these criteria. Because of that fact, various pharmaceutical technologies, such as CDs (Loftsson & Järvinen, 1999) and prodrugs (Järvinen & Järvinen, 1996), have been applied to improve the physicochemical properties of ophthalmic drugs. They are also useful approaches for the development of ophthalmic cannabinoids.

In addition to the lipophilicity and aqueous solubility of a drug, molecular size (Liaw & Robinson, 1992), charge (Liaw et al., 1992), and degree of ionization (Maren & Jankowska, 1985; Brechue & Maren, 1993) also affect corneal absorption. Tear fluid has a limited buffering capacity (Carney & Hill, 1979). Thus, pH and buffering capacity of the instilled solution affect the pH of the tear fluid and, consequently, drug ionization on the pre-corneal area. The non-ionized form of the drug usually penetrates the cornea more easily than the ionized form, so the pH and buffering capacity of an instilled solution can have a significant effect on ophthalmic drug absorption.

3. Cyclodextrins for ophthalmic drug delivery

3.1. Structure and function

CDs are macrocyclic oligosaccharides containing D-(+)-glucopyranose subunits joined through α -1,4-bonds (Fig. 2a). The most common natural CDs are α -CD, β -CD, and γ -CD, which contain six, seven, and eight glucose molecules, respectively. Each CD has varying inclusion capabilities that stem from differences in their internal cavity dimensions (Stella et al., 1999; Thompson, 1997).



 $R = H \Rightarrow \beta$ -cyclodextrin $R \neq H \Rightarrow \beta$ -cyclodextrin derivative

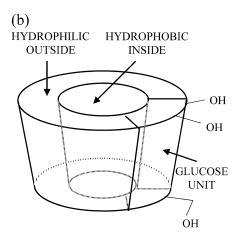


Fig. 2. Chemical structure (a) and the molecular shape (b) of β -CD.

CDs are cone-shaped molecules that are open at both ends (Fig. 2b). All their hydrophilic hydroxy groups are located on the outside of this structure, and the inside is relatively hydrophobic. These external OH groups can be chemically altered to produce CD derivatives with modified properties. Various CD derivatives have been developed to increase the aqueous solubility and safety of natural CDs, especially β -CD. Currently, the CD derivatives that are expected to have commercial pharmaceutic utility are randomly methylated derivatives of β -CD, 2-hydroxypropyl derivatives of β -CD (HP- β -CD), and a sulfobutylether derivative of β -CD (SBE- β -CD) (Thompson, 1997; Szente & Szejtli, 1999).

The most important feature of a CD is its ability to act as a "host" in the formation of inclusion complexes with hydrophobic "guest" molecules. An inclusion complex is formed when the guest molecule, or part of this molecule, enters into the hydrophobic cavity of the host CD. This complex is not a static entity, but a dynamic relationship between host and guest, and consists of the free drug, the drug/CD complex, and the unoccupied CD in an equilibrium that is dependent on several factors (Frömming & Szejtli, 1994; Szejtli, 1988). Complex formation depends on the

size, shape, and polarity of the guest molecule and on the properties of the CD that is used. Complexation can be used to increase the aqueous solubility, dissolution rate, stability, and bioavailability of drugs, as well as to modify the local irritation potential and organoleptic properties of therapeutic agents (Loftsson, 1995; Loftsson & Brewster, 1996; Rajewski & Stella, 1996; Zhang & Rees, 1999).

The safety profiles of natural CDs and their most commonly used derivatives have been reviewed recently (Irie & Uekama, 1997; Thompson, 1997). In general, natural CDs and most derivatives are unable to penetrate biological membranes. Thus, orally administered CDs are practically nontoxic. Furthermore, safety evaluations have shown that at least γ -CD, HP- β -CD, and SBE- β -CD appear to be safe, even when administered parenterally. However, α -CD, β -CD, and the methylated CDs are not suitable for parenteral administration.

3.2. Ophthalmic applications

In ophthalmology, CDs mainly have been used to improve the delivery of poorly water-soluble drugs (Loftsson & Järvinen, 1999; Loftsson & Stefansson, 1997). As pointed out by Rajewski and Stella (1996), poorly water-soluble drugs are frequently administered as suspensions or ointment-dosage forms, which may cause eye irritation and blurred vision, respectively. In addition, this limited water solubility hinders drug dissolution into tear fluid on the precorneal area and results in poor ophthalmic bioavailability (Bary et al., 2000; Fridriksdottir et al., 1997; Kristinsson et al., 1996).

Ophthalmic administration of CDs reveals some basic differences compared with the use of CDs via other routes of drug administration (Järvinen, K. et al., 1995; Rajewski & Stella, 1996). As it is generally assumed that only the free drug, and not the drug/CD complex, can penetrate across

biological barriers, its release from this inclusion complex must take place before absorption can occur (Nakanishi et al., 1989; Frijlink et al., 1990). In contrast to oral and parenteral administration, ophthalmic preparations do not exhibit a significant increase in the fraction of free drug available after ocular administration. This is due to a lack of drug/CD-complex dilution by the small tear fluid volume $(\approx 7 \,\mu\text{L})$. In addition, aqueous eyedrops are removed from the pre-corneal area within a few minutes (Chrai et al., 1973), and, thus, the drug/CD complex may not have enough time to release an effective dose of the drug before its clearance from the pre-corneal area. Especially in ophthalmic applications, excessive complexation of a drug may decrease its bioavailability (Bary et al., 2000; Davies et al., 1997; Järvinen, T. et al., 1995; Reer et al., 1994), although this drawback may be overcome by increasing the viscosity of the aqueous eyedrop formulation (Jarho et al., 1996a). Fig. 3 shows the kinetic parameters assumed to be critical for ophthalmic preparations containing CDs.

Studies dealing with the use of CDs in ophthalmic formulations usually involve in vitro (rarely in vivo) absorption studies. The in vitro studies are usually performed by monitoring the flux of a drug through semi-permeable membranes (Loftsson et al., 1994b) or the isolated corneas of experimental animals (Bary et al., 2000; Kearse & Green, 2000; Siefert et al., 1999). Both in vitro and in vivo studies have shown that the complexation of poorly water-soluble drugs with CDs increases corneal penetration/absorption of the drug compared with a suspension of the compound or a control preparation (Loftsson et al., 1994a; Reer et al., 1994; Siefert et al., 1999). This improved penetration/absorption is due to a fast dissociation of the drug/CD complex in solution, compared with the slow dissolution of a solid-drug suspension.

The interaction of CDs with cell membranes is a potential mechanism for increasing the bioavailability of ocular drugs

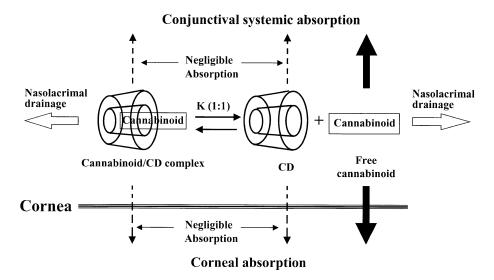


Fig. 3. Basic kinetic processes assumed to be critical in ophthalmic drug delivery in the presence of CDs.

(Reddy et al., 1996). In ophthalmic drug delivery, the most commonly studied CDs (HP- β -CD and SBE- β -CD) do not cause damage to corneal tissues (Jansen et al., 1990; Järvinen et al., 1994). Although pretreatment of the cornea with HP- β -CD and SBE- β -CD does not affect in vitro corneal penetration of water-soluble pilocarpine (Järvinen et al., 1994; Jarho et al., 1997), HP- β -CD (Freedman et al., 1993) and α -CD (Keipert et al., 1996) have been reported to increase its miotic effect. In addition, α -CD has been shown to improve the corneal penetration of pilocarpine due to CD-induced membrane effects (Siefert & Keipert, 1997). Methylated CDs are not commonly used in ophthalmic studies due to their damaging effect on the cornea (Jansen et al., 1990).

3.3. Cannabinoid administration

Cannabinoids have very poor aqueous solubility, which hinders their use in topical ophthalmic preparations. CDs are able to increase, by several orders of magnitude, the aqueous solubility of either AEA and its derivatives (Jarho et al., 1996b) or Δ^9 -THC (Jarho et al., 1998). In addition, CDs significantly improve the aqueous stability of AEA (Jarho et al., 1996b). Thus, CDs would seem ideal as potential pharmaceutical excipients for the formulation of ophthalmic cannabinoids.

Fig. 4 shows how increasing CD concentrations affect the transcorneal flux of AEA solutions. With AEA mostly in suspension, its flux increases with greater CD solution strength, making an increase of AEA complexation possible. However, when all the AEA molecules have already been dissolved, this flux decreases at still greater CD concentrations due to excess drug/CD complexation and thus, a decreased concentration of free AEA in the donor phase. This study, and other similar studies through skin (Sigurdardottir & Loftsson, 1995) or nasal mucosa (Kublik

et al., 1996), demonstrates the importance of an optimal CD concentration for ophthalmic applications. Recently, Kearse and Green (2000) reported the effect of various vehicles upon the in vitro transcorneal penetration of Δ^9 -THC. Various vehicles were compared with the light mineral oil generally studied as a vehicle for topical Δ^9 -THC applications, and α -CD showed significantly enhanced flux and penetration values. Interestingly, β -CD, γ -CD, and HP- β -CD did not significantly enhance flux and permeability compared with light mineral oil. However, the employed concentrations of γ -CD (20%) and HP- β -CD (30%) were too high, which may explain their modest flux values. The enhanced Δ^9 -THC transcorneal flux shown with the α -CD vehicle is most probably due to the corneal effects of the latter (Siefert & Keipert, 1997).

4. Ocular cannabinoid systems

4.1. Endogenous cannabinoids

Two endogenous cannabinoid receptor ligands, AEA and 2-arachidonoylglycerol (2-AG) have been isolated from both nervous and peripheral tissues (Devane et al., 1992b; Mechoulam et al., 1995; Sugiura et al., 1995; Felder et al., 1996). The endogenous cannabinoids mimic the actions of Δ^9 -THC, but are inactivated rapidly in vivo or in vitro. The inactivation of AEA or 2-AG occurs through a carrier-mediated cellular uptake process, followed by enzymatic hydrolysis, in cultured brain neurons and astrocytoma cells (Di Marzo et al., 1994; Beltramo & Piomelli, 2000; Bisogno et al., 2001). In cells, AEA is hydrolyzed to arachidonic acid and ethanolamine by fatty acid amide hydrolase (FAAH) (Deutsch & Chin, 1993; Cravatt et al., 1996; Ueda et al., 1995). The enzymatic hydrolysis of 2-AG to arachidonic acid and glycerol presumably is catalyzed by a yet unchar-

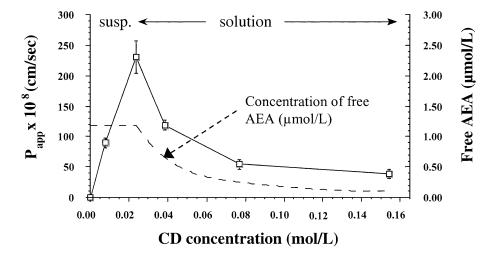


Fig. 4. Permeability (P_{app} , mean \pm S.E., n = 2 - 6) of AEA through the isolated rabbit cornea as a function of HP- β -CD concentration (solid line) and calculated concentration of free AEA on the donor side as a function of HP- β -CD concentration (broken line). susp., suspension. Modified from Jarho et al. (1996b).

acterized monoacylglycerol lipase activity (Beltramo & Piomelli, 2000). Di Marzo et al. (1998b) have suggested that FAAH may be one of the enzymes deputed to the physiological inactivation of 2-AG. However, the contribution of FAAH to hydrolysis of 2-AG seems to be minor (Goparaju et al., 1999). The properties and the possible physiological roles of AEA and 2-AG have been discussed comprehensively in earlier reviews (Mechoulam et al., 1998; Piomelli et al., 2000; Bisogno et al., 2001).

The hydrolysis and biosynthesis of AEA in various porcine ocular tissues, including retina, iris, choroid, lacrimal gland, and optic nerve, has been reported (Matsuda et al., 1997). Both AEA synthase activity (1.9-4.2 nmol/ min⁻¹/mg⁻¹ protein at 37°C) and hydrolase activity $(1.2-3.5 \text{ nmol/min}^{-1}/\text{mg}^{-1} \text{ protein at } 37^{\circ}\text{C})$ in the ocular tissues were comparable with those of brain homogenate. The highest enzyme activity was in the retina. However, condensation of AEA from arachidonic acid and ethanolamine under physiological conditions is unlikely, as the required arachidonic acid and ethanolamine concentrations are high (for recombinant rat liver FAAH, K_m values are 190 µM and 36 mM for arachidonic acid and ethanolamine, respectively) (Ueda et al., 1995; Kurahashi et al., 1997). Another biosynthetic pathway for AEA through phosphodiesterase-mediated cleavage of N-arachidonoylphosphatidylethanolamine has been suggested in cultured brain neurons (Di Marzo et al., 1994). The bovine retina has been shown to contain AEA and N-docosahexanoylethanolamine and their putative direct biosynthetic precursors, N-arachidonoylphosphatidylethanolamine and N-docosahexanoylphosphatidylethanolamine (Bisogno et al., 1999). In addition, FAAH-like activity that is inhibitable by the FAAH inhibitors phenylmethylsulfonyl fluoride (PMSF) and arachidonoyltrifluoromethylketone has also been identified in bovine retina (Bisogno et al., 1999), and an FAAH enzyme protein has been localized from rat retina using immunocytochemical methods (Yazulla et al., 1999).

Both endogenous cannabinoids AEA and 2-AG have been identified from bovine and rat retinas (Bisogno et al., 1999; Straiker et al., 1999b). The amounts of AEA and 2-AG in the bovine retina determined by gas chromatography-electron impact mass spectrometry were 64.0 ± 9.6 pmol/g and 1.63 ± 0.31 nmol/g of retinal tissue, respectively (Bisogno et al., 1999). The bovine retina contained 25 times more 2-AG than AEA. In rat retina, 2-AG (2.97 ±0.066 nmol/g) was found in amounts similar to that of the brain (Straiker et al., 1999b), but AEA was not detectable. Other endogenous cannabinoid-like lipids, such as *N*-palmitoyle-thanolamide (a CB₂ receptor agonist) and *N*-oleoylethanolamine, were also identified from rat retina (Straiker et al., 1999b).

4.2. Cannabinoid receptors

Two cannabinoid receptor subtypes have been characterized from human and animal tissues (Devane et al., 1988;

Munro et al., 1993). The CB₁ receptor is distributed throughout the CNS, whereas the CB₂ receptor is localized predominantly in peripheral tissues. Both cannabinoid receptors belong to the G-protein-coupled receptor superfamily.

The first indirect evidence for the possible existence of ocular cannabinoid receptors was provided in 1996 when Schlicker et al. inhibited dopamine release in guinea pig retinal discs by an application of the CB₁ receptor agonists WIN-55,212-2 and CP-55,940. The effect was reversed with the CB₁ receptor antagonist SR 141716A. This study was followed by the observation that metabolically stable forms of AEA (Pate et al., 1997, 1998) and CP-55,940 (Pate et al., 1998) lowered IOP in rabbits, an effect that was eliminated for either type compound upon subcutaneous pretreatment of the animals with SR 141716A (Pate et al., 1998). Subsequently, CB₁ receptor mRNA was identified in various ocular tissues of the rat (Porcella et al., 1998). CB₁ receptor mRNA is more abundant in the ciliary body area and iris than in the retina and choroid. In the human eye, most of the CB₁ receptor mRNA also appears to be in the ciliary body (Porcella et al., 2000).

By the use of subtype-specific affinity-purified polyclonal antibodies against the CB₁ receptor protein, a wide distribution of CB₁ receptors has been determined within the human anterior eye and retina (Straiker et al., 1999a). CB₁ receptors are present in the human ciliary epithelium, corneal epithelium and endothelium, trabecular meshwork, Canal of Schlemm, ciliary muscle, and in blood vessels of the ciliary body. In addition, retinal tissues of humans (Straiker et al., 1999a) and of several animal species (Straiker et al., 1999b; Yazulla et al., 1999, 2000) have been shown to contain CB₁ receptors. Evidence for the expression of CB₂ receptor mRNA has been obtained from rat retinal tissues by the use of in situ hybridization histochemistry and reverse transcription polymerase chain reaction (Lu et al., 2000).

The wide existence of cannabinoid receptors (Table 1) and their endogenous ligands in various eye tissues suggests a physiological role for the cannabinoid system in various ocular functions (Porcella et al., 1998; Straiker et al., 1999a; Lu et al., 2000). Endogenous cannabinoids acting via cannabinoid receptors may have effect, for example, on aqueous humor production and outflow, as well as on vision itself. Identification of the ocular cannabinoid system components enables the development of novel drugs that act specifically via the cannabinoid receptors of the eye.

5. Cannabinoids and intraocular pressure

Cannabinoids are generally classified according to their various chemical structures into the following four main groups: (1) classical cannabinoids (i.e., phytocannabinoids and synthetic congeners), (2) nonclassical cannabinoids (e.g., bicyclic compounds or nitrogen isosters), (3) amino-

Table 1
Regional distribution of cannabinoid receptors in the eye

Eye tissue	CB_1	CB ₂	Species	Reference					
Retina	X, Y Y		Human, rat, mouse, monkey, goldfish, chick	Straiker et al., 1999a, 1999b; Yazulla et al., 1999, 2000; Lu et al., 2000; Porcella et al., 2000					
Ganglion cell layer, inner plexiform layer, inner nuclear layer, outer plexiform layer, photoreceptors									
Cornea Epithelium, endothelium	X		Human	Straiker et al., 1999a					
Iris	X, Y		Human, rat	Porcella et al., 1998, 2000; Straiker et al., 1999a					
Trabecular meshwork	X		Human	Straiker et al., 1999a					
Schlemm's Canal	X		Human	Straiker et al., 1999a					
Ciliary Body	X, Y		Human, rat	Porcella et al., 1998, 2000; Straiker et al., 1999a					
Non-pigmented epithelium, muscle fibers									
Choroid	Y		Rat	Porcella et al., 1998					

X, expression of cannabinoid receptor protein; Y, expression of cannabinoid receptor mRNA.

alkylindoles, and (4) eicosanoids (i.e., endocannabinoid congeners and related enzyme inhibitors). The chemical structures of prototypic members of each group are shown in Figs. 5-7.

5.1. Classical cannabinoids

Classical cannabinoids retain the various natural cannabinoid ring structures with their oxygen atoms. Typical of this class are the phytocannabinoids Δ^9 -THC, cannabidiol, and cannabinol and the commercial synthetic drugs syn-

Fig. 5. Chemical structures of typical classical cannabinoids.

hexyl and nabilone (Fig. 5). Hepler and Frank (1971) first published that smoked marijuana reduces IOP. As a result, several animal and human studies involving systemic and topical administration of Δ^9 -THC or other classical cannabinoids have been carried out. Although HU-211 belongs to this taxon, its IOP effects, as discussed in Section 5.5, are not due to CB₁ receptor interactions.

5.1.1. Systemic administration

Intravenous (ElSohly et al., 1981, 1984; Green et al., 1983) and oral (Howes, 1984; Merritt et al., 1980b; Waller et al., 1984) administration of various cannabinoids, including Δ^9 -THC, Δ^8 -THC, nabilone, and cannabinol, reduces IOP in animals and humans. However, it soon became apparent from the human studies that systemic routes of administration carried the burden of such undesirable side-effects as psychoactivity, conjunctival hyperemia, and postural hypotension (Green & McDonald, 1987), and, thus, more effort has been focused on studies with ophthalmic administration.

5.1.2. Ophthalmic administration

Topical application of cannabinoids to the eye seems to be the obvious alternative for minimizing systemic drug concentrations and maximizing the dose at the site of action. However, the hydrophobic cannabinoids have remarkably poor water solubility (Garrett & Hunt, 1974), so investigations employing several more physically compatible vehicles have been made, including sesame oil (Green & Bowman, 1976; Green et al., 1977, 1978) or mineral oil (Green et al., 1977, 1978; Merritt et al., 1981; Jay & Green, 1983), the latter of which worked best. Unfortunately, this vehicle can be irritating (Jay & Green, 1983) and has been demonstrated to influence IOP itself (Merritt et al., 1986).

Fig. 6. Chemical structure of CP-55,940 (nonclassical cannabinoid) and WIN-55,212-2 (aminoalkylindole).

Colasanti et al. (1984a, 1984b, 1984c) employed polyethylene glycol for acute doses, as well as chronic administration via osmotic pumps. Use of cannabinoids in aqueous media with the aid of Tween 80 (Green et al., 1977), submicron aqueous emulsification (Muchtar et al., 1992), and CD complexes (Arsenovic, 1997; Jarho et al., 1998) have yielded various degrees of success.

Such research reveals that certain cannabinoids, after topical administration, are more efficacious than others, but that there remains a large degree of variability between these studies (Colasanti, 1986; Green, 1998, 2000). Much of this difference can be blamed upon vehicle choice and the fact that ocular absorption is very poor (Green et al., 1977) and probably inconsistent. The vast majority of drug not locally absorbed is available for systemic circulation (Chiang et al., 1983), and may affect other parts of the body.

5.2. Nonclassical cannabinoids

Nonclassical cannabinoids include bicyclic analogs of Δ^9 -THC that lack a pyran "B" ring (Johnson & Melvin, 1986). The most important member of this group is CP-55,940 (Fig. 6), a compound that is broadly used in cannabinoid receptor-binding studies (Pertwee, 1997).

The topical application of CP-55,940 significantly decreases IOP in normotensive rabbits (Pate et al., 1998) and in those with elevated IOP (Sugrue et al., 1996). The unilateral ocular administration of CP-55,940 did not cause a decrease of IOP in untreated eyes equal to that of treated eyes, which suggests that the locus of action may be the eye rather than the CNS. The IOP decrease was neutralized by a subcutaneous pretreatment with SR 141716 (2.5 mg/kg), which suggests CB₁ receptor involvement with local IOP effects (Pate et al., 1998).

5.3. Aminoalkylindoles

Aminoalkylindoles form one important class of cannabinoids, the prototype molecule being WIN-55,212-2 (Fig. 6). WIN-55,212-2 binds to both the CB₁ and CB₂ receptors, but has a several-fold bias toward the CB₂ receptor (Felder & Glass, 1998; Pertwee, 1997; Showalter et al., 1996) and exhibits cannabinoid-like activity both in vitro and in vivo

(Compton et al., 1992). WIN-55,212-3, the enantiomer of WIN-55,212-2, is inactive in both cannabinoid receptor systems (Compton et al., 1992).

Sugrue et al. (1996) first reported that topical administration of WIN-55,212-2 significantly decreases IOP in rabbits and monkeys, but that WIN-55,212-3 is significantly less active. The study also showed that a topical dose of WIN-55,212-2 decreased aqueous humor inflow 66%, whereas outflow was unchanged in the rabbits.

Hodges et al. (1997) reported that intravenous injection of WIN-55,212-2 (3 mg/kg) and of another cannabinoid did not cause a statistically significant IOP decrease in rabbits. However, they did mention that observed IOPs tended to decrease more substantially in some animals than in others.

Song and Slowey (2000) demonstrated that topical application of WIN-55,212-2 (dissolved in HP- β -CD) significantly decreased IOP in the treated eyes of rabbits, but no significant IOP reduction was observed in contralateral eyes. The maximal IOP reduction by 100 μ g of WIN-55,212-2 was 4.7±0.5 mm Hg at 2 hr after topical application. A topical dose (25 μ g) of the CB₁ receptor antagonist SR141716A significantly attenuated the IOP effect of 100- μ g WIN-55,212-2, which supports the results of Pate et al. (1998) and their hypothesis that a CB₁ receptor is involved in the IOP reduction effects of cannabinoid agonists. The inactive WIN-55,212-3 enantiomer did not decrease IOP.

Recently, Porcella et al. (2001) reported that a topical dose of WIN-55,212-2 (25 or 50 μg dissolved in HP- β -CD) significantly decreased the IOP of human glaucoma patients

Fig. 7. Chemical structures of typical endocannabinoids and their congeners.

whose symptoms were refractory to conventional glaucoma therapy.

5.4. Endocannabinoids and their congeners

5.4.1. Arachidonylethanolamide and 2-arachidonoylglycerol Topical administration of AEA (Fig. 7) (Mikawa et al., 1997; Pate et al., 1995) and 2-AG (Fig. 7) (Pate et al., 1996) to normotensive rabbits caused an initial increase and subsequent decrease in IOP in treated eyes. In untreated eyes, only a very weak IOP decrease (Pate et al., 1995) or no IOP decrease (Mikawa et al., 1997) was observed. The maximum hypotensive effect occurred 1 or 2 hr after topical administration.

Subcutaneous indomethacin prevented the IOP effects of AEA (Pate et al., 1996). Indomethacin is a cyclo-oxygenase (COX) inhibitor, and prevents prostaglandin synthesis from arachidonic acid (Fig. 8). Earlier it had been suggested that it is a prostaglandin rather than arachidonic acid itself that affects IOP after arachidonic acid treatment (Podos et al., 1973a, 1973b). Thus, it seems that topically administered AEA is catabolized (Fig. 8) in the eye to arachidonic acid (see Section 4), which is subsequently biosynthesized to prostaglandins responsible for the IOP effects. Subcutaneous pretreatment with the FAAH inhibitor PMSF eliminated the

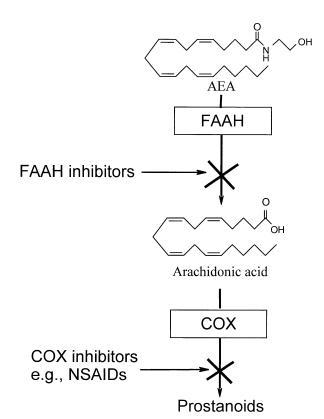


Fig. 8. Catabolism of AEA to arachidonic acid and subsequent biosynthesis of prostanoids.

typical initial increase of IOP observed in AEA-treated eyes, although a significant hypotension was apparent (Laine et al., 2002). When the experiment was repeated with a subcutaneous co-injection of SR 141716A (a highly specific CB₁ receptor antagonist), this hypotensive effect was eliminated, suggesting that IOP reduction caused by the apparently undergraded AEA may be mediated via a CB₁ receptor.

5.4.2. α-Substituted anandamides

Topically administered α -substituted anandamides, such as α -methyl-AEA (Fig. 7) and α -isopropyl-AEA, lacked an initial increase of IOP, but caused immediate ocular hypotension in the treated eye (Pate et al., 1997). In the untreated eye, a significant IOP decrease was usually not observed. Subcutaneous injection (2.5 mg/kg) of SR 141716A eliminated the IOP reduction caused by topically administered α-isopropyl-AEA (Pate et al., 1998), which suggests that these IOP effects are mediated via a CB₁ receptor. Subcutaneous administration of SR 141716A alone elevated the IOP of rabbits, which may indicate that it is acting either as a competitive antagonist of endogenous AEA or as an inverse CB₁ receptor agonist (Bouaboula et al., 1997). In contrast, a topical AEA positive control, apparently working via a prostanoid mechanism, was unaffected by subcutaneously pre-administered SR 141716A, which indicates that the prevention of α -isopropyl-AEA effects by SR 141716A probably was not due to the sum of unrelated effects.

5.4.3. Anandamide uptake inhibitors

An interesting approach by Laine et al. (2001) for lowering IOP via a putative CB₁ mechanism was inferred from the works of Beltramo et al. (1997), Calignano et al. (1997), and Di Marzo et al. (1998a), and employed endocannabinoid transport inhibitors (Fig. 7) to increase levels of endogenous ocular AEA or 2-AG. They reported that a topical dose of AM404 (62.5 μg) administered in HP-β-CD significantly decreased IOP in rabbits. However, if AM404 was given in propylene glycol, a significant IOP increase (without subsequent IOP decrease) was observed. The authors concluded that the latter IOP increase may be due to a greater absorption of AM404 by the eye, followed by its degradation to arachidonic acid. In contrast, topical administration of olvanil (312.5 µg) in propylene glycol caused a significant IOP reduction, without provoking an initial hypertensive phase. This was thought most probably due to the fact that olvanil is a derivative of oleic acid (Fig. 7), which does not serve as a substrate for prostaglandin synthesis.

AM404 and olvanil activates vanilloid receptor Type 1 (VR1) (De Petrocellis et al., 2000). Consequently, it can be argued that the IOP effects of AM404 and olvanil might be mediated via VR1 receptors. However, it is not currently known if VR1 receptors are present in the eye, and if so, whether or not they are involved in regulating IOP. The IOP effects of selective endocannabinoid uptake inhibitors, such

as VDM11 and VDM13 (De Petrocellis et al., 2000), would be useful to study in order to eliminate the possible IOP effects via VR1 receptors.

5.5. Bimatoprost

Bimatoprost (Fig. 7) is a highly efficacious and longacting ocular hypotensive agent (Woodward et al., 2001). It mimics the activity of a newly discovered family of fatty acid amides, termed "prostamides," that may be biosynthesized from AEA. Bimatoprost exhibits no activity on CB_1 or CB_2 receptors, which suggests that its IOP effects cannot be ascribed to an interaction with or stimulation of cannabinoid receptors. Bimatoprost is not classified as a cannabinoid due to its prostanoid chemical structure and lack of CB receptor activity.

5.6. HU-211

HU-211 (Fig. 5) is a non-psychoactive synthetic cannabinoid, although its enantiomer (HU-210) is one of the most potent cannabinoids described thus far (Devane et al., 1992a). HU-211 does not appreciably bind to the CB₁ receptor (Howlett et al., 1990), which explains this lack of cannabimimetic activity (Mechoulam et al., 1988). HU-211 is currently under evaluation as a commercial drug candidate for preventing the secondary biochemical injury that is collateral to head trauma or brain inflammation, probably via a *N*-methyl-D-aspartate (NMDA) receptor mechanism (Feigenbaum et al., 1989; Love, 1999).

Intravenous administration of HU-211 results in a dose-related IOP decrease in rabbits that is stronger than that of Δ^9 -THC or Δ^8 -THC (Beilin et al., 2000). This HU-211 effect can be attenuated by pretreatment with yohimbine (an α_2 -adrenergic antagonist) and propranolol (a β -adrenergic antagonist).

A topical dose of HU-211 (0.12% as a submicron emulsion) significantly decreased IOP in normotensive rabbits (Naveh et al., 2000). The maximal IOP reduction was 5.3 ± 1.8 mm Hg at 1.5 hr after drug administration, and a statistically significant IOP decrease was also observed at 4 and 6 hr after drug administration. In the contralateral eye, a statistically significant IOP decrease was observed only at 4 hr after drug administration.

6. Mechanisms of action

6.1. Intraocular pressure reduction

The actual mechanism of action for cannabinoid reduction of IOP is unknown. Because smoking marijuana reduces blood pressure (Crawford & Merritt, 1979; Merritt et al., 1980a), the obvious hypothesis is that IOP reduction is simply a reflection of this change. Although systemic hypotension may have such an ocular influence, it probably

cannot account for most of the observed IOP effect (Korczyn, 1980).

Until recently, the influence of ophthalmic cannabinoids on IOP has been assumed to be mediated through the CNS rather than locally. However, a substantial difference in IOP between the cannabinoid treated versus untreated eyes of cats (Colasanti et al., 1984a, 1984b, 1984c) supports the local effect hypothesis. Systemic absorption of the vast majority of a topically applied dose (Chiang et al., 1983) via blood circulation (Chang & Lee, 1987; Urtti & Salminen, 1993) and subsequent transfer of the drug to the untreated eye (Salminen & Urtti, 1984) may account for minor contralateral effects. Although the concentration of a drug in the untreated eye may be substantially lower than that of the treated eye, these lower concentrations are often sufficient to cause some reduction of IOP (Urtti & Salminen, 1985). This bilateral IOP asymmetry might also be explained as a combination of major localized and minor CNS effects. However, direct administration of THC into the cerebral ventricles of rabbits, or ventriculocisternal perfusion, does not affect IOP (Liu & Dacus, 1987).

Recent studies using the CB₁ receptor antagonist SR141716A have implied that the IOP reduction caused by cannabinoids is mediated via CB₁ receptors. Topically applied AEA (Pate et al., 1995) and other α -unsubstituted anandamides (Pate et al., 1996) seem to influence IOP through their hydrolysis to arachidonic acid, which is a COX pathway precursor of the prostanoids. In contrast, metabolically stable α -substituted anandamides (Pate et al., 1997) and other types of cannabinoids, e.g., CP-55,940 (Pate et al., 1998) and WIN-55,212-2 (Song & Slowey, 2000), seem to act via CB₁ receptors. However, simultaneous administration of the FAAH inhibitor PMSF apparently prevents degradation of exogenous AEA (Laine et al., 2002), as reflected by a disappearance of the typical initial hypertension. This apparently intact AEA acts upon a CB₁ receptor, as evidenced by the fact that its hypotensive effects could be eliminated via the use of SR141716A. An overall scheme for IOP physiology, including AEA metabolism, theorized from the studies published to date is presented in Fig. 9.

The anatomical distribution of ocular cannabinoid receptors (Straiker et al., 1999a) indicates that endogenous cannabinoids may have a physiological role for the regulation of ocular pressure. The existence of CB₁ receptors in the trabecular meshwork and in the Canal of Schlemm suggests a possible influence of cannabinoids on conventional aqueous humor outflow. CB₁ receptors of the ciliary pigment epithelium and ciliary muscle imply an effect on either (or both) aqueous humor production and uveoscleral outflow. Additional mechanistic studies are required to determine possible involvement of the sympathetic or parasympathetic nervous systems, and any possible vascular component (Kaufman & Wis, 1998).

The non-psychoactive synthetic cannabinoid HU-211 apparently reduces IOP (Naveh et al., 2000; Beilin et al.,

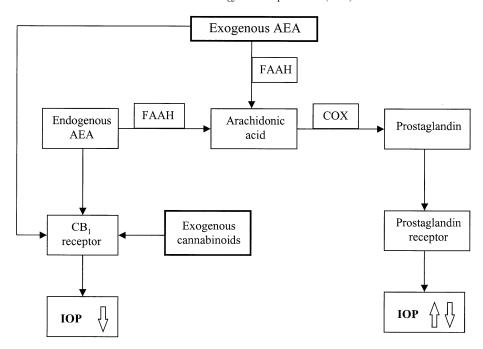


Fig. 9. The proposed ocular AEA metabolism and IOP dynamics.

2000), despite its relative inactivity at the CB₁ receptor (Howlett et al., 1990). Its mechanism of action is unknown at present. An attempt to counter this effect by the use of a CB receptor antagonist has not been attempted yet. HU-211 is an NMDA receptor antagonist (Feigenbaum et al., 1989; Nadler et al., 1993), and to our knowledge, evidence correlating NMDA receptor antagonism to a decrease in IOP has yet to be published.

6.2. Neurological implications

Elevated IOP is only one factor in the pathophysiology of glaucoma, and its influence on the retinal disk is indirect. Indeed, a significant percentage of glaucoma patients exhibit normotensive IOP (Sommer et al., 1991; Collaborative Normal Tension Glaucoma Study Group, 1998a, 1998b). At present, it is commonly accepted that glaucoma is a degenerative disease of the optic nerve (Schwartz & Yoles, 2000). Retinal disc damage is thought to occur through at least two mechanisms: occlusion of axonal flow, which causes localized interference with delivery of organelles (e.g., Hochmann & Herkenham, 1999) and cytosolic factors (Nickells, 1999), and restriction of the microcirculation that nourishes the optic nerve (Prunte et al., 1998). While both phenomena are influenced by IOP, the latter may also occur independently.

On the intracellular level, retinal disk ischemia causes anoxia and hypoglycemia, resulting in neuronal toxicity and ultimately in apoptosis (Nickells, 1999; Nickells & Zack, 1996). This, in turn, contributes to a toxic extracellular environment that may result in an ongoing progression of optic nerve degeneration (Schwartz & Yoles, 1999). The secondary neuronal injury observed in "stroke" victims acts

by a similar mechanism (Love, 1999). Among the products produced by this process are an excess of neurotransmitters, such as glutamate (Duarte et al., 1998; Sucher et al., 1997) and nitric oxide (Adachi et al., 1998a, 1998b). The former compound induces a neuronal "hyperexcitability" of the NMDA receptor and results in a large flux of Ca²⁺ (Pauwels et al., 1991; Ritch, 2000; Schousboe et al., 1997) and other ions (Pauwels et al., 1991; Yu et al., 1999) across the membrane, which induces cell death. Excessive quantities of nitric oxide synergize apoptosis (Leist et al., 1997) and act as an oxidative free radical initiator, which induces a cascade of other destructive effects, including lipid membrane peroxidation. Tumor necrosis factor- α has been suggested (Shohami et al., 1997) to be a primary mediator of neurotoxicity after brain trauma, and its possible involvement in retinal ischemia also bears scrutiny.

Although the effect of HU-211 on vascular tissues is unknown, it shares many of the neurotherapeutic effects of THC. It has been investigated for use in both secondary brain (Leker et al., 1999) and optic nerve (Yoles et al., 1996; Schwartz & Yoles, 1999) injury. Its actions include a combination of effects: open channel blockade of the NMDA Ca²⁺ channel (Eshhar et al., 1993; Nadler et al., 1993), non-glutamate/glycine receptor antagonism (Feigenbaum et al., 1989), tumor necrosis factor-α inhibition (Shohami et al., 1997), nitric oxide suppression (Gallily et al., 1997), and free radical scavenging (Biegon & Joseph, 1995). Ca²⁺-channel blockers have shown promise for clinical use in glaucoma (Netland et al., 1993), and the ability of HU-211 to also antagonize NMDA receptor activity is complementary. Interestingly, AEA has demonstrated an ability to modulate this receptor (Hampson et al.,

1998b), and palmitoylethanolamide has also been shown to exhibit an indirect anti-excitotoxic effect involving a CB₂-like receptor (Skaper et al., 1996).

The overall implication for the sum of these studies is potentially significant. Cannabinoids may reveal themselves to be useful for the treatment of glaucoma in a quite comprehensive manner: lowering IOP, restoring microcirculation, inhibiting apoptosis, and minimizing free radical damage, among other mechanisms. This hypothetical combination would supersede that employed by any currently employed glaucoma drug, and may help to explain why the smoking of marijuana has preserved the sight of those unresponsive to other glaucoma therapies.

7. Cannabinoids for glaucoma therapy

It is well documented that various cannabinoids are able to reduce IOP when administered orally, intravenously, or by inhalation (e.g., Green, 1998). Generally, it is thought that the clinical use of cannabinoids in the treatment of glaucoma is hindered by difficulties with preparing appropriate ophthalmic dosage forms and because of their potential for psychoactive side-effects. Novel drug delivery technologies, such as CDs (Jarho et al., 1996b, 1998) or microemulsions (Muchtar et al., 1992), have enabled preparation of topically administered cannabinoids that decrease IOP. This is a clear improvement over lipid-based vehicles such as the oils used in earlier studies of topical cannabinoids. The question of CNS side-effects is a more open question, although moot if the total delivered dose is less than that needed to elicit psychoactivity. The following approaches may provide the means by which an ophthalmic cannabinoid pharmaceutical can be developed.

7.1. Optimal drug delivery properties

Only a few percent of an ophthalmic dose is delivered to the intraocular tissues. The major part of this dose will be absorbed into the systemic circulation, which often leads to side-effects (Fig. 1). If ocular absorption can be improved by formulation approaches or by molecular modification, the therapeutic goal can be achieved by a smaller topical dose, with consequent reduction of undesired effects. This is an especially important consideration in the development of ophthalmic medicines affecting CB₁ receptors. The pro-drug strategy may also improve the physicochemical and biopharmaceutical properties of ophthalmic cannabinoids (Järvinen & Järvinen, 1996), and has been applied successfully to the development of other glaucoma drugs, e.g., dipivefrin (Mandell et al., 1978) and latanoprost (Resul et al., 1993). Recently, O-1057 was reported as a possible water-soluble prodrug of THC derivation (Pertwee et al., 2000) and anandamide phosphates as potential water-soluble prodrugs of anandamides (Järvinen et al., 2001).

7.2. CB₂ receptor approach

Recent IOP (Pate et al., 1998; Laine et al., 2002; Song & Slowey, 2000) and receptor (Straiker et al., 1999a, 1999b; Yazulla et al., 1999, 2000) studies strongly suggest that IOP effects of cannabinoids are mediated via ocular CB₁ receptors. However, studies should be carried out to determine if populations of CB₂ receptors exist on anatomically relevant areas of the eye and if CB₂-specific receptor agonists or antagonists have an IOP effect.

7.3. Cannabinoid "soft-drugs"

A "soft-drug" is designed to undergo rapid metabolic deactivation after affecting its target area. An example of ophthalmic application of this drug strategy is loteprednol etabonate, an active corticosteroid that lacks the usual steroidal systemic side-effects (Bodor, 2000). An ideal "soft" ophthalmic cannabinoid would be absorbed by the eye and have a pharmacological effect, but would become inactivated in the systemic circulation, thus limiting its effect on target tissues. Several soft cannabinoids are currently under development (Buchwald et al., 2000).

After topical administration, AEA could potentially act as an ophthalmic soft-drug to decrease IOP via an ocular CB_1 receptor. However the catabolism of its amide bond leading to the formation of CB_1 -inactive arachidonic acid is actually too rapid and results in an elevation of IOP. Adding an α -carbon substituent protects against this metabolic vulnerability, defaulting the molecule to a slower metabolism at other structural locations (e.g., double bonds) and allowing CB_1 activity in the eye.

8. Conclusions

Rapid advances in cannabinoid research have been achieved in less than a decade. These characterizations have provided basic tools for a closer look at the mechanisms behind the well-known efficacy of marijuana on glaucoma. The recent demonstration of endocannabinoid IOP mechanisms and the subsequent discovery of CB₁ receptors on appropriate intraocular structures suggests a physiological role for these compounds in the regulation of normal ocular tension, and implies a route by which a new class of anandamide-based glaucoma drugs may be developed. Topical delivery of these or other cannabinoids directly to the site of action should eliminate psychoactivity, but suffer practical problems of administration, which can be overcome through modern techniques of molecular design and drug delivery. The prospect of cannabinoid glaucoma medicines promises someday to substitute simple IOP reduction for a more comprehensive package of ocular therapeutic support.

Acknowledgements

This work was supported by the Academy of Finland and The National Technology Agency of Finland.

References

- Adachi, K., Fujita, Y., Morizane, C., Akaike, A., Ueda, M., Satoh, M., Masai, H., Kashii, S., & Honda, Y. (1998a). Inhibition of NMDA receptors and nitric oxide synthase reduces ischemic injury of the retina. *Eur J Pharmacol* 350, 53–57.
- Adachi, K., Kashii, S., Masai, H., Ueda, M., Morizane, C., Kaneda, K., Kume, T., Akaike, A., & Honda, Y. (1998b). Mechanism of the pathogenesis of glutamate neurotoxicity in retinal aschemia. *Graefes Arch Clin Exp Ophthalmol* 236, 766–774.
- Ahmed, I., Gokhale, R. D., Shah, M. V., & Patton, T. F. (1987). Physico-chemical determinants of drug diffusion across the conjunctiva, sclera, and cornea. *J Pharm Sci* 76, 583–586.
- Alward, W. L. M. (1998). Medical management of glaucoma. *Engl J Med* 339, 1298–1307.
- Arsenovic, J. (1997). Formulation of aqueous opthalmic solutions of cannabinoids for use in glaucoma therapy. M.Ph. Thesis, University of Otago.
- Bary, A. R., Tucker, I. G., & Davies, N. M. (2000). Considerations in the use of hydroxypropyl-β-cyclodextrin in the formulation of aqueous ophthalmic solutions of hydrocortisone. *Eur J Pharm Biopharm 50*, 237–244.
- Beilin, M., Neumann, R., Belkin, M., Green, K., & Bar-Ilan, A. (2000). Pharmacology of the intraocular pressure (IOP) lowering effect of systemic dexanabinol (HU-211), a non-psychotropic cannabinoid. *J Ocul Pharmacol Ther* 16, 217–230.
- Beltramo, M., & Piomelli, D. (2000). Carrier-mediated transport and enzymatic hydrolysis of the endogenous cannabinoid 2-arachidonoyl glycerol. Neuroreport 11, 1231–1235.
- Beltramo, M., Stella, N., Calignano, A., Lini, S. Y., Makriannis, A., & Piomelli, D. (1997). Functional role of high-affinity anandamide transport, as revealed by selective inhibition. *Science* 277, 1094–1096.
- Biegon, A., & Joseph, A. B. (1995). Development of HU-211 as a neuroprotectant for ischemic brain damage. *Neurol Res* 17, 275–280.
- Bisogno, T., Delton-Vandenbrouke, I., Milone, A., Lagarde, M., & Di Marzo, V. (1999). Biosynthesis and inactivation of N-arachidonoylethanolamine (anandamide) and N-docosahexaenoylethanolamine in bovine retina. Arch Biochem Biophys 370, 300–307.
- Bisogno, T., Maccarrone, M., De Petrocellis, L., Jarrahian, A., Finazzi-Agro, A., Hillard, C., & Di Marzo, V. (2001). The uptake by cells of 2-arachidonoylglycerol, an endogenous agonist of cannabinoid receptors. *Eur J Biochem* 268, 1982–1989.
- Bodor, N. (2000). Recent advances in retrometabolic drug design and targeting approaches. *Pharmazie* 55, 163–166.
- Borchardt, R. T. (1990). Assessment of transport barriers using cell and tissue culture systems. *Drug Dev Ind Pharm 16*, 2595–2612.
- Bouaboula, M., Perrachon, S., Milligan, L., Canat, X., Rinaldi-Carmona, M., Portier, M., Barth, F., Calandra, B., Pecceu, F., Lupker, J., Maffrand, J. P., Le Fur, G., & Casellas, P. (1997). A selective inverse agonist for central cannabinoid receptor inhibits mitogen-activated protein kinase activation stimulated by insulin or insulin-like growth factor 1. Evidence for a new model of receptor/ligand interactions. *J Biol Chem* 272, 22330–22339.
- Brechue, W. F., & Maren, T. H. (1993). pH and drug ionization affects ocular pressure lowering of topical carbonic anhydrase inhibitors. *Invest Ophthalmol Vis Sci* 34, 2581–2587.
- Buchwald, A., Browne, C. E., Wu, W.-M., & Bodor, N. (2000). Soft cannabinoid analogues as potential anti-glaucoma agents. *Pharmazie* 55, 196–201.

- Calignano, A., La Rana, G., Beltramo, M., Makriyannis, A., & Piomelli, D. (1997). Potentiation of anandamide hypotension by the transport inhibitor, AM404. Eur J Pharmacol 337, R1-R2.
- Carney, L. G., & Hill, R. M. (1979). Human tear buffering capacity. Arch Ophthalmol 97, 951–952.
- Chang, S. C., & Lee, V. H. L. (1987). Nasal and conjunctival contributions to the systemic absorption of topical timolol in the pigmented rabbit: implications in the design of strategies to maximize the ratio of ocular to systemic absorption. *J Ocul Pharmacol* 3, 159–169.
- Chiang, C.-W., Barnett, G., & Brine, D. (1983). Systemic absorption of Δ⁹-tetrahydrocannabinol after ophthalmic administration to the rabbit. J Pharm Sci 72, 136–138.
- Chien, D.-S., Sasaki, H., Bundgaard, H., Buur, A., & Lee, V. H. L. (1991).
 Role of enzymatic lability in the corneal and conjunctival penetration of timolol ester prodrugs in the pigmented rabbits. *Pharm Res* 8, 728–733.
- Chrai, S. S., Patton, T. F., Mehta, A., & Robinson, J. R. (1973). Lacrimal and instilled fluid dynamics in rabbit eye. J Pharm Sci 62, 1112–1121.
- Colasanti, B. K. (1986). Ocular hypotensive effect of marihuana cannabinoids: correlate of central action or separate phenomenon? *J Ocul Phar*macol 2, 295–304.
- Colasanti, B. K., Brown, R. E., & Craig, C. R. (1984a). Ocular hypotension, ocular toxicity, and neurotoxicity in response to marijuana extract and cannabidiol. *Gen Pharmacol* 15, 479–484.
- Colasanti, B. K., Craig, C. R., & Allara, R. D. (1984b). Intraocular pressure, ocular toxicity and neurotoxicity after administration of cannabinol or cannabigerol. *Exp Eye Res* 39, 251–259.
- Colasanti, B. K., Powell, S. R., & Craig, C. R. (1984c). Intraocular pressure, ocular toxicity and neurotoxicity after administration of D9-tetra-hydrocannabinol or cannabichromene. Exp Eye Res 38, 63-71.
- Collaborative Normal Tension Glaucoma Study Group (1998a). Comparison of glaucomatous progression between untreated patients with normal-tension glaucoma and and patients with therapeutically reduced introcular pressures. Am J Ophthalmol 126, 487–497.
- Collaborative Normal Tension Glaucoma Study Group (1998b). The effectiveness of intraocular pressure reduction in the treatment of normal-tension glaucoma. *Am J Ophthalmol* 126, 498–505.
- Compton, D. R., Gold, L. H., Ward, S. J., Balster, R. L., & Martin, B. R. (1992). Aminoalkylindole analogs: cannabimimetic activity of a class of compounds structurally distinct from Δ⁹-tetrahydrocannabinol. *J Pharmacol Exp Ther 263*, 1118–1126.
- Cooler, P., & Gregg, J. M. (1977). The effect of delta-9-tetrahydrocannabinol on intraocular pressure in humans. South Med J 70, 951–954.
- Cravatt, B. F., Giang, D. K., Mayfield, S. P., Boger, D. L., Lerner, R. A., & Gilula, N. B. (1996). Molecular characterization of an enzyme that degrades neuromodulatory fatty-acid amides. *Nature* 384, 83–87.
- Crawford, W. J., & Merritt, J. C. (1979). Effect of tetrahydrocannabinol on arterial and intraocular hypertension. *Int J Clin Pharmacol Biopharm* 17, 191–196.
- Davies, N. M. (2000). Biopharmaceutical considerations in topical ocular drug delivery. Clin Exp Pharmacol Physiol 27, 558–562.
- Davies, N. M., Wang, G., & Tucker, I. G. (1997). Evaluation of a hydrocortisone/hydroxypropyl-β-cyclodextrin solution for ocular drug delivery. Int J Pharm 156, 201–209.
- De Petrocellis, L., Bisogno, T., Davis, J. B., Pertwee, R. G., & Di Marzo, V. (2000). Overlap between the ligand recognition properties of the anandamide transporter and VR1 vanilloid receptor: inhibitors of anandamide uptake with negligible capsaicin-like activity. FEBS Lett 483, 52–56.
- Deutsch, D. G., & Chin, S. A. (1993). Enzymatic synthesis and degradation of anandamide, a cannabinoid receptor agonist. *Biochem Pharmacol* 46, 791–796.
- Devane, W. A., Dysarz, I. F. A., Johnson, M. R., Melvin, L. S., & Howlett, A. C. (1988). Determination and characterization of a cannabinoid receptor in rat brain. *Mol Pharmacol* 34, 605-613.
- Devane, W. A., Breuer, A., Sheskin, T., Jarbe, T. U. C., Eisen, M. S., & Mechoulam, R. (1992a). A novel probe for the cannabinoid receptor. J Med Chem 35, 2065–2069.

- Devane, W. A., Hanus, L., Breuer, A., Pertwee, R. G., Stevenson, L. A., Griffin, G., Gibson, D., Mandelbaum, A., Etinger, A., & Mechoulam, R. (1992b). Isolation and structure of a brain constituent that binds to the cannabinoid receptor. *Science* 258, 1946–1949.
- Di Marzo, V., Fontana, A., Cadas, H., Schinelli, S., Cimino, C., Schwartz, J.-C., & Piomelli, D. (1994). Formation and inactivation of endogenous cannabinoid anandamide in central neurons. *Nature* 372, 686–691.
- Di Marzo, V., Bisogno, T., Melck, D., Ross, R., Brockie, H., Stevenson, L., Perwee, R., & De Petrocellis, L. (1998a). Interactions between synthetic vanilloids and the endogenous cannabinoid system. FEBS Lett 436, 449–454.
- Di Marzo, V., Bisogno, T., Sugiura, T., Melck, D., & De Petrocellis, L. (1998b). The novel endogenous cannabinoid 2-arachidonoylglycerol is inactivated by neuronal- and basophil-like cells: connections with anandamide. Biochem J 331, 15-19.
- Doane, M. G., Jensen, A. D., & Dohlman, C. H. (1978). Penetration routes of topically applied eye medication. Am J Ophthalmol 85, 383–386.
- Duarte, C. B., Ferreira, I. L., Santos, P. F., Carvalho, A. L., Agostinho, P. M., & Carvalho, A. P. (1998). Glutamate in life and death of retinal amacrine cells. *Gen Pharmacol* 30, 289–295.
- ElSohly, M. A., Harland, E. C., Murphy, J. C., Wirth, P., & Waller, C. W. (1981). Cannabinoids in glaucoma: a primary screening procedure. J Clin Pharmacol 21, 472S–478S.
- ElSohly, M. A., Harland, E. C., Benigni, D. A., & Waller, C. W. (1984).
 Cannabinoids in glaucoma II: the effect of different cannabinoids on intraocular pressure of the rabbit. Curr Eye Res 3, 841–850.
- Eshhar, N., Striem, S., & Biegon, A. (1993). HU-211, a non-psychotropic cannabinoid, rescues cortical neurones from excitatory amino acid toxicity in culture. *Neuroreport* 5, 237–240.
- Feigenbaum, J. J., Bergmann, F., Richmond, S. A., Mechoulam, R., Nadler, V., Kloog, Y., & Sokolovsky, M. (1989). A non-psychotropic cannabinoid acts as a functional N-methyl-D-aspartate (NMDA) receptor blocker. Proc Natl Acad Sci USA 86, 9584–9587.
- Felder, C. C., & Glass, M. (1998). Cannabinoid receptors and their endogenous agonists. *Annu Rev Pharmacol Toxicol* 38, 179–200.
- Felder, C. C., Nielsen, A., Briley, E. M., Palkovits, M., Priller, J., Axelrod, J., Nguyen, D. N., Richardson, J. M., Riggin, R. M., Koppel, G. A., Paul, S. M., & Becker, G. W. (1996). Isolation and measurement of the endogenous cannabinoid receptor agonist, anandamide, in brain and peripheral tissues of human and rat. FEBS Lett 393, 321–325.
- Freedman, K. A., Klein, J. W., & Crosson, C. E. (1993). Beta-cyclodextrins enhanced biovailability of pilocarpine. *Curr Eve Res* 12, 641–647.
- Fridriksdottir, H., Loftsson, T., & Stefansson, E. (1997). Formulation and testing of methazolamide cyclodextrin eye drop solutions. *J Controlled Release* 44, 95–99.
- Frijlink, H. W., Eissens, A. C., Schoonen, A. J. M., & Lerk, C. F. (1990). The effect of cyclodextrins on drug absorption. II. In vivo observation. *Int J Pharm 64*, 195–205.
- Frömming, K.-H., & Szejtli, J. (1994). Cyclodextrins in Pharmacy. Dordrecht: Kluwer Academic Publishers.
- Gallily, R., Yamin, A., Waksmann, Y., Ovadia, H., Weidenfeld, J., Bar-Joseph, A., Biegon, A., Mechoulam, R., & Shohami, E. (1997). Protection against septic shock and suppression of tumor necrosis factor alpha and nitric oxide production by dexanabinol (HU-211), a nonpsychotropic cannabinoid. J Pharmacol Exp Ther 283, 918–924.
- Garrett, E. R., & Hunt, C. A. (1974). Physico-chemical properties, solubility and protein binding of *delta-9*-tetrahydrocannabinol. *J Pharm Sci* 63, 1056–1064.
- Goparaju, S. K., Ueda, N., Taniguchi, K., & Yamamoto, S. (1999). Enzymes of porcine brain hydrolyzing 2-arachidonoylglycerol, an endogenous ligand of cannabinoid receptors. *Biochem Pharmacol* 57, 417–423
- Green, K. (1998). Marijuana smoking vs cannabinoids for glaucoma therapy. Arch Ophthalmol 116, 1433–1437.
- Green, K. (2000). Marihuana and intraocular pressure. In: G. G. Nahas, K. N. Sutin, & S. Agurell (Eds.), *Marihuana and Medicine* (pp. 581–589). Totawa: Humana Press Inc.

- Green, K., & Bowman, K. (1976). Effect of marihuana and derivatives on aqueous humor dynamics in the rabbit. In Braude, M. C., & Szara, S. (Eds.), *The Pharmacology of Marihuana*, Vol 2 (pp. 803–813). New York: Raven Press.
- Green, K., & Mcdonald, T. F. (1987). Ocular toxicology of marijuana: an update. J Toxicol Cutaneous Ocul Toxicol 6, 309–334.
- Green, K., Bigger, J. F., Kim, K., & Bowman, K. (1977). Cannabinoid penetration and chronic effects in the eye. *Exp Eye Res* 24, 197–205.
- Green, K., Wynn, H., & Bowman, K. A. (1978). A comparison of topical cannabinoids on intraocular pressure. *Exp Eye Res* 27, 239–246.
- Green, K., Symonds, M. C., Oliver, N. W., & Elijah, R. D. (1983). Intraocular pressure following systemic administration of cannabinoids. *Curr Eye Res* 2, 247–253.
- Hampson, A. J., Bornheim, L. M., Scanziani, M., Yost, C. S., Gray, A. T., Hansen, B. M., Leonoudakis, D. J., & Bickler, P. E. (1998). Dual effects of anandamide on NMDA receptor-mediated responses and neurotransmission. *J Neurochem* 70, 671–676.
- Hepler, R. S., & Frank, I. M. (1971). Marihuana smoking and intraocular pressure. JAMA 217, 1392.
- Hochmann, A. G., & Herkenham, M. (1999). Cannabinoid receptors undergo axonal flow in sensory nerves. *Neuroscience* 92, 1171–1175.
- Hodges, L. C., Reggio, P. H., & Green, K. (1997). Evidence against cannabinoid receptor involvement in intraocular pressure effects of cannabinoids in rabbits. *Ophthalmol Res* 29, 1–5.
- Howes, J. F. (1984). Antiglaucoma effects of topically and orally administered cannabinoids. In: Agurell S., Dewey, W. L., & Willette, R. E. (Eds.), *The Cannabinoids: Chemical, Pharmacologic and Therapeutic Aspects* (pp. 881–890). New York: Academic Press Inc.
- Howlett, A. C., Champion, T. M., Wilken, G. H., & Mechoulam, R. (1990). Stereochemical effects of 11-OH-D-8-tetrahydrocannabinol-dimethyl-heptyl to inhibit adenylate cyclase and bind to the cannabinoid receptor. Neuropharmacology 29, 161–165.
- Huang, H.-S., Schoenwald, R. D., & Lach, J. L. (1983). Corneal penetration behavior of beta-blocking agents II: assessment of barrier contributions. *J Pharm Sci* 72, 1272–1286.
- Irie, T., & Uekama, K. (1997). Pharmaceutical applications of cyclodextrins. III. Toxicological issues and safety evaluation. J Pharm Sci 86, 147–162.
- Jansen, T., Xhonneux, B., Mensensm, J., & Borgers, M. (1990). Beta-cyclodextrins as vehicles in eye-drop formulations: an evaluation of their effects on rabbit corneal epithelium. Lens Eye Toxic Res 7, 459–468.
- Jarho, P., Järvinen, K., Urtti, A., Stella, V., & Järvinen, T. (1996a). Modified β-cyclodextrin (SBE7-β-CyD) with viscous vehicle improves the ocular delivery and tolerability of pilocarpine prodrug in rabbits. *J Pharm Pharmacol* 48, 263–269.
- Jarho, P., Urtti, A., Pate, D. W., Suhonen, P., & Järvinen, T. (1996b). Increase in aqueous solubility, stability and in vitro corneal permeability of anandamide by hydroxypropyl-β-cyclodextrin. *Int J Pharm 137*, 209–217.
- Jarho, P., Järvinen, K., Urtti, A., Stella, V. J., & Järvinen, T. (1997). The effect of SBE7-β-cyclodextrin with viscous vehicle on ocular delivery of pilocarpine prodrug in rabbits. In: Szejtli, J., & Szente, L. (Eds.), Proceedings of the Eigth International Symposium on Cyclodextrins (pp. 299–402). Dordrecht: Kluwer Academic Publishers.
- Jarho, P., Pate, D. W., Brenneisen, R., & Järvinen, T. (1998). Hydroxypropyl- β -cyclodextrin and its combination with hydroxylpropyl-methylcellulose increases aqueous solubility of Δ^9 -tetrahydrocannabinol. *Life Sci* 63, PL381–PL384.
- Järvinen, K., Järvinen, T., Thompson, D. O., & Stella, V. J. (1994). The effect of a modified β-cyclodextrin, SBE4-β-CD, on the aqueous stability and ocular absorption of pilocarpine. Curr Eye Res 13, 897–905.
- Järvinen, K., Järvinen, T., & Urtti, A. (1995). Ocular absorption following topical delivery. Adv Drug Delivery Rev 16, 3-19.
- Järvinen, T., & Järvinen, K. (1996). Prodrugs for improved ocular drug delivery. Adv Drug Delivery Rev 19, 203–224.
- Järvinen, T., Järvinen, K., Urtti, A., Thompson, D. O., & Stella, V. J. (1995). Sulfobutyl ether β-cyclodextrin (SBE4-β-CD) in eyedrops im-

- proves the tolerability of a topically applied pilocarpine prodrug in rabbits. *J Ocul Pharmacol Ther 11*, 95–106.
- Järvinen, T., Juntunen, J., Huuskonen, J., Nevalainen, T., Pate, D. W., & Laine, K. (2001). Water-soluble prodrugs of anandamides. In Abstracts of the International Cannabinoid Research Society Meeting, June 28–30, 2001, San Lorenzo de El Escorial, Spain (p. 15). Burlington: International Cannabinoid Research Society.
- Jay, W. M., & Green, K. (1983). Multiple-drop study of topically applied $1\% \Delta^9$ -tetrahydrocannabinol in human eyes. *Arch Opthalmol 101*, 591–593.
- Johnson, M. R., & Melvin, L. S. (1986). The discovery of nonclassical cannabinoid analgetics. In Mechoulam, R. (Ed.), Cannabinoids as Therapeutic Agents (pp. 121–145). Boca Raton: CRC Press.
- Kaufman, P. L., & Wis, M. (1998). Marijuana and glaucoma. Arch Ophthalmol 116, 1512–1513.
- Kearse, E. C., & Green, K. (2000). Effect of vehicle upon in vitro transcorneal permeability and intracorneal content of Δ^9 -tetrahydrocannabinol. *Curr Eye Res 20*, 496–501.
- Keipert, S., Fedder, J., Böhm, A., & Hanke, B. (1996). Interactions between cyclodextrins and pilocarpine—as an example of a hydrophilic drug. *Int J Pharm 142*, 153–162.
- Korczyn, A. D. (1980). The ocular effects of cannabinoids. Gen Pharmacol 11, 419–423.
- Kristinsson, J. K., Fridriksdottir, H., Thorisdottir, S., Sigurdardottir, A. M., Stefansson, E., & Loftsson, T. (1996). Dexamethasone-cyclodextrinpolymer co-complexes in aqueous eye drops. *Invest Ophthalmol Vis* Sci 37, 1199–1203.
- Kublik, H., Bock, T. K., Schreier, H., & Müller, B. W. (1996). Nasal absorption of 17β-estradiol from different cyclodextrin inclusion formulations in sheep. Eur J Pharm Biopharm 42, 320–324.
- Kurahashi, Y., Ueda, N., Suzuki, N., Suzuki, M., & Yamamoto, S. (1997). Reversible hydrolysis and synthesis of anandamide demonstrated by recombinant rat fatty-acid amide hydrolysis. *Biochem Biophys Res Commun* 237, 512–515.
- Laine, K., Järvinen, T., Savinainen, J., Laitinen, J. T., Pate, D. W., & Järvinen, K. (2001). Effects of topical anandamide uptake inhibitors, AM404 and olvanil, on intraocular pressure in normotensive rabbits. *Pharm Res* 18, 494–499.
- Laine, K., Järvinen, K., Pate, D. W., Urtti, A., & Järvinen, T. (2002). Effect of the enzyme inhibitor, phenylmethylsulfonyl fluoride, on the IOP profiles of topical anandamides. *Invest Ophthalmol Vis Sci* 43, 393–397.
- Leist, M., Fava, E., Montecucco, C., & Nicotera, P. (1997). Peroxynitrite and nitric acid donors induce neuronal apoptosis by eliciting autocrine excitotoxicity. *Eur J Neurosci 9*, 1488–1498.
- Leker, R. R., Shohami, E., Abramsky, O., & Ovadia, H. (1999). Dexanabinol; a novel neuroprotective drug in experimental focal cerebral ischemia. *J Neurol Sci* 162, 114–119.
- Liaw, J., & Robinson, J. R. (1992). The effect of polyethylene glycol molecular weight on corneal transport and the related influence of penetration enhancers. *Int J Pharm* 88, 125–140.
- Liaw, J., Rojanasakul, Y., & Robinson, J. R. (1992). The effect of drug charge type and charge density on corneal transport. *Int J Pharm 88*, 111–124.
- Liu, J. H. K., & Dacus, A. C. (1987). Central nervous system and peripheral mechanisms in ocular hypotensive effect of cannabinoids. *Arch Oph-thalmol* 105, 245–258.
- Loftsson, T. (1995). Effects of cyclodextrins on the chemical stability of drugs in aqueous solutions. *Drug Stability 1*, 22–33.
- Loftsson, T., & Brewster, M. E. (1996). Pharmaceutical applications of cyclodextrins. 1. Drug solubilization and stabilization. *J Pharm Sci* 85, 1017–1025.
- Loftsson, T., & Järvinen, T. (1999). Cyclodextrins in ophthalmic drug delivery. Adv Drug Delivery Rev 36, 59-79.
- Loftsson, T., & Stefansson, E. (1997). Effect of cyclodextrins on topical drug delivery to the eye. *Drug Dev Ind Pharm* 23, 473–481.
- Loftsson, T., Fridriksdottir, H., Thorisdottir, S., & Stefansson, E. (1994a).

- The effect of hydroxypropyl methylcellulose on release of dexamethazone from aqueous 2-hydroxypropyl- β -cyclodextrin formulations. *Int J Pharm 104*, 181–184.
- Loftsson, T., Fridriksdottir, H., Thorisdottir, S., Stefansson, E., Sigurdardottir, A. M., Gudmundsson, Ö., & Sigthorsson, T. (1994b). 2-Hydroxy-propyl-β-cyclodextrinin topical carbonic anhydrase inhibitor formations. *Eur J Pharm Sci 1*, 175–180.
- Love, S. (1999). Oxidative stress in brain ischemia. *Brain Pathol 9*, 119–131.
- Lu, Q., Straiker, A., Lu, G., & Maguire, G. (2000). Expression of CB2 receptor mRNA in adult rat retina. Vis Neurosci 17, 91–95.
- Mandell, A. I., Stentz, F., & Kitabchi, A. E. (1978). Dipivalyl epinephrine: a new prodrug in the treatment of glaucoma. *Ophthalmology* 85, 268–275.
- Maren, T. H., & Jankowska, L. (1985). Ocular pharmacology of sulfonamides: the cornea as barrier and depot. Curr Eye Res 4, 399–408.
- Matsuda, S., Kanemitsu, N., Nakamura, A., Mimura, Y., Ueda, N., Kurahashi, Y., & Yamamoto, S. (1997). Metabolism of anandamide, an endogenous cannabinoid receptor ligand, in porcine ocular tissues. *Exp Eye Res* 64, 707-711.
- Mechoulam, R., Feigenbaum, J. J., Lander, N., Segal, M., Jarbe, T. U. C., Hiltunen, A. J., & Consroe, P. (1988). Enantiomeric cannabinoids: stereospecificity of psychotropic activity. *Experientia* 44, 762–764.
- Mechoulam, R., Ben-Shabat, S., Hanus, L., Ligumsky, M., Kaminski, N. E.,
 Schatz, A. R., Gopher, A., Almog, S., Martin, B. R., Compton, D. R.,
 Pertwee, R. G., Griffin, G., Bayewitch, M., Barg, J., & Vogel, Z. (1995).
 Identification of an endogenous 2-monoglyceride, present in canine gut,
 that binds to cannabinoid receptors. *Biochem Pharmacol* 50, 83–90.
- Mechoulam, R., Fride, E., & Di Marzo, V. (1998). Endocannabinoids. Eur J Pharmacol 359, 1–18.
- Merritt, J. C., Crawfford, W. J., Alexander, P. C., Anduze, A. L., & Gelbart, S. S. (1980a). Effect of marihuana on intraocular and blood pressure in glaucoma. *Ophthalmology* 877, 222–228.
- Merritt, J. C., McKinnon, S., Armstrong, J. R., Hatem, G., & Reid, L. A. (1980b). Oral Δ^9 -tetrahydrocannabinol in heterogenous glaucomas. *Ann Ophthalmol* 12, 947–950.
- Merritt, J. C., Perry, D. D., Russell, D. N., & Jones, B. F. (1981). Topical Δ⁹-tetrahydrocannabinol and aqueous dynamics in glaucoma. *J Clin Pharmacol* 21, 471S–476S.
- Merritt, J. C., Shrewsbury, R. P., Locklear, F., Demby, K. B., & Wittle, G. L. (1986). Effects of Δ⁹-tetrahydrocannabinol and vehicle constituents on intraocular pressure in normotensive dogs. *Res Comm Subst Abuse* 7, 29–35
- Mikawa, Y., Matsuda, S., Kanagawa, T., Tajika, T., Ueda, N., & Mimura, Y. (1997). Ocular activity of topically administered anandamide in the rabbit. *Jpn J Ophthalmol* 41, 217–220.
- Muchtar, S., Almog, S., Torracca, M. T., Saettone, M. F., & Benita, S. (1992). A sub-micron emulsion as ocular vehicle for *delta-8*-tetrahy-drocannabinol: effect on intraocular pressure in rabbits. *Ophthalmic Res* 24, 142–149.
- Munro, S., Thomas, K. L., & Abu-Shaar, M. (1993). Molecular characterization of a peripheral receptor for cannabinoids. *Nature* 365, 61-65.
- Nadler, V., Mechoulam, R., & Sokolovsky, M. (1993). Blockade of Ca²⁺ influx through the N-methyl-D-aspartate receptor ion channel by the non-psychoactive cannabinoid HU-211. Brain Res 622, 79–85.
- Nakanishi, K., Masada, M., Nadai, T., & Miyajima, K. (1989). Effect of the interaction of drug-β-cyclodextrin complex with bile salts on the drug absorption from rat small intestinal lumen. Chem Pharm Bull 37, 211–214.
- Naveh, N., Weissman, C., Muchtar, S., Benita, S., & Mechoulam, R. (2000). A submicron emulsion of HU-211, a synthetic cannabinoid, reduces intraocular pressure in rabbits. *Graefes Arch Exp Ophthalmol* 238, 334–339.
- Netland, P. A., Chaturvedi, N., & Dreyer, E. B. (1993). Calcium channel blockers in the management of low-tension and open-angle glaucoma. *Am J Ophthalmol* 115, 608–613.

- Nickells, R. W. (1999). Apoptosis of retinal ganglion cells in glaucoma: an update of the molecular pathways involved in cell death. Surv Ophthalmol 43, S151–S161.
- Nickells, R. W., & Zack, D. J. (1996). Apoptosis in ocular disease: a molecular overview. Ophthalmic Paediatr Genet 17, 145–165.
- Pate, D. W., Järvinen, K., Urtti, A., Jarho, P., & Järvinen, T. (1995). Ophthalmic arachidonyl ethanolamide decreases intraocular pressure in normotensive rabbits. *Curr Eye Res* 14, 791–797.
- Pate, D. W., Järvinen, K., Urtti, A., Jarho, P., Fich, M., Mahadevan, V., & Järvinen, T. (1996). Effects of topical anandamides on intraocular pressure in normotensive rabbits. *Life Sci* 58, 1849–1860.
- Pate, D. W., Järvinen, K., Urtti, A., Jarho, P., Mahadevan, V., & Järvinen, T. (1997). Effects of topical alpha-substituted anandamides on intraocular pressure in normotensive rabbits. *Pharm Res* 14, 1738–1743.
- Pate, D. W., Järvinen, K., Urtti, A., Mahadevan, V., & Järvinen, T. (1998). Effect of the CB1 receptor antagonist, SR 141716A, on cannabinoid-induced ocular hypotension in normotensive rabbits. *Life Sci 63*, 2181–2188.
- Pauwels, P. J., Leysen, J. E., & Janssen, P. A. (1991). Ca⁺⁺ and Na⁺ channels involved in neuronal cell death. Protection by flunarizine. *Life Sci* 48, 1881–1893.
- Pertwee, R. G. (1997). Pharmacology of cannabinoid CB1 and CB2 receptors. *Pharmacol Ther* 74, 129–180.
- Pertwee, R. G., Gibson, T. M., Stevenson, L. A., Ross, R. A., Banner, W. K., Saha, B., Razdan, R. K., & Martin, B. R. (2000). O-1057, a potent watersoluble cannabinoid receptor agonist with antinociceptive properties. *Br J Pharmacol* 129, 1577–1584.
- Piomelli, D., Giuffrida, A., Calignano, A., & de Fonseca, F. R. (2000). The endocannabinoid system as a target for therapeutic drugs. *Trends Phar-macol Sci* 21, 218–224.
- Podos, S. M., Becker, B., & Kass, M. A. (1973a). Prostaglandin synthesis, inhibition, and intraocular pressure. *Invest Ophthalmol* 12, 426–433.
- Podos, S. M., Becker, B., & Kass, M. A. (1973b). Indomethacin blocks arachidonic acid-induced elevation of intraocular pressure. *Prostaglan-dins* 3, 7–16.
- Porcella, A., Casellas, P., Gessa, G. L., & Pani, L. (1998). Cannabinoid receptor CB1 mRNA is highly expressed in the rat ciliary body: implications for the antiglaucoma properties of marihuana. *Mol Brain Res* 58, 240–245.
- Porcella, A., Maxia, C., Gessa, G. L., & Pani, L. (2000). The human eye expresses high levels of CB1 cannabinoid receptor mRNA and protein. *Eur J Neurosci* 12, 1123–1127.
- Porcella, A., Maxia, C., Gessa, G. L., & Pani, L. (2001). The synthetic cannabinoid WIN55212-2 decreases the intraocular pressure in human glaucoma resistant to conventional therapies. *Eur J Neurosci* 13, 409–412.
- Prunte, C., Orgul, S., & Flammer, J. (1998). Abnormalities of microcirculation in glaucoma: facts and hints. Curr Opin Ophthalmol 9, 50-55.
- Rajewski, R. A., & Stella, V. J. (1996). Pharmaceutical applications of cyclodextrins. 2. In vivo drug delivery. J Pharm Sci 85, 1142–1169.
- Reddy, I. K., Khan, M. A., Wu, W.-M., & Bodor, N. S. (1996). Permeability of a soft steroid loteprednol etabonate, through an excised rabbit cornea. *J Ocul Pharmacol Ther 12*, 159–167.
- Reer, O., Bock, T. K., & Müller, B. W. (1994). In vitro corneal permeability of diclofenac sodium in formulations containing cyclodextrins compared to commercial product voltaren ophtha. *J Pharm Sci* 83, 1345–1349.
- Resul, B., Stjernschantz, J., No, K., Liljebris, C., Selen, G., Astin, M., & Bito, L. Z. (1993). Phenyl-substituted prostaglandins: potent and selective antiglaucoma agents. J Med Chem 36, 243–248.
- Ritch, R. (2000). Neuroprotection: is it already applicable to glaucoma therapy. Curr Opin Ophthalmol 11, 78–84.
- Rojanasakul, Y., Wang, L.-Y., Bhat, M., Glover, D. D., Malanga, C. J., & Ma, J. K. H. (1992). The transport barrier of epithelia: a comparative study on membrane permeability and charge selectivity in the rabbit. *Pharm Res* 9, 1029–1034.
- Salminen, L., & Urtti, A. (1984). Disposition of ophthalmic timolol in

- treated and untreated rabbit eyes. A multiple and single dose study. Exp Eve Res 38, 203-206.
- Sasaki, H., Yamamura, K., Mukai, T., Nishida, K., Nakamura, J., Nakashima, M., & Ichikawa, M. (1999). Enhancement of ocular drug penetration. Crit Rev Ther Drug Carrier Syst 16, 85–146.
- Schlicker, E., Timm, J., & Göthert, M. (1996). Cannabinoid receptor-mediated inhibition of dopamine release in the retina. *Naunyn Schmiedebergs Arch Pharmacol* 354, 791–795.
- Schoenwald, R. D., & Huang, H.-S. (1983). Corneal penetration behaviour of β-blocking agents. I: physicochemical factors. *J Pharm Sci* 72, 1266–1272.
- Schoenwald, R. D., & Ward, R. (1978). Relationship between steroid permeability across excited rabbit cornea and octanol-water partition coefficients. J Pharm Sci 67, 786–788.
- Schousboe, A., Belhage, B., & Frandsen, A. (1997). Role of Ca²⁺ and other second messengers in excitatory amino acid receptor mediated neurodegeneration: clinical perspectives. *Clin Neurosci* 4, 194–198.
- Schwartz, M., & Yoles, E. (1999). Optic nerve degeneration and potential neuroprotection: implications for glaucoma. Eur J Ophthalmol 9, S9–S11.
- Schwartz, M., & Yoles, E. (2000). Neuroprotection: a new treatment modality for glaucoma. Curr Opin Ophthalmol 11, 107–111.
- Shohami, E., Gallily, R., Mechoulam, R., Bass, R., & Ben-Hur, T. (1997). Cytokine production in the brain following closed head injury: dexanabinol (HU-211) is a novel TNF-alpha inhibitor and an effective neuro-protectant. J Neuroimmunol 72, 169–177.
- Showalter, V. M., Compton, D. R., Martin, B. R., & Abood, M. E. (1996). Evaluation of binding in a transfected cell line expressing a peripheral cannabinoid receptor (CB2): identification of cannabinoid receptor subtype selective ligands. *J Pharmacol Exp Ther* 278, 989–999.
- Siefert, B., & Keipert, S. (1997). Influence of alpha-cyclodextrin and hydroxylated β-cyclodextrin derivatives on the in vitro corneal uptake and permeation of aqueous pilocarpine-HCL solutions. J Pharm Sci 86, 716–720.
- Siefert, B., Pleyer, U., Muller, M., Hartmann, C., & Keipert, S. (1999). Influence of cyclodextrins on the in vitro corneal permeability and in vivo ocular distribution of thalidomide. *J Ocul Pharmacol Ther 15*, 429–438.
- Sigurdardottir, A. M., & Loftsson, T. (1995). The effect of polyvinylpyrrolidone on cyclodextrin complexation of hydrocortisone and its diffusion through hairless mouse skin. *Int J Pharm 126*, 73–78.
- Skaper, S. D., Buriani, A., Dal-Toso, R., Petrelli, L., Romanello, S., Facci, L., & Leon, A. (1996). The ALIAmide palmitoylethanolamide and cannabinoids, but not anandamide, are protective in a delayed postglutamate paradigm of excitotoxic death in cerebellar granule neurons. *Proc Natl Acad Sci USA 93*, 3984–3989.
- Sommer, A., Tielsch, J. M., Katz, J., Quigley, H. A., Gottsch, J. D., Javitt, J., & Singh, K. (1991). Relationship between intraocular pressure and primary open angle glaucoma among white and black Americans. *Arch Ophthalmol* 109, 1090–1095.
- Song, Z.-H., & Slowey, C.-A. (2000). Involvement of cannabinoid receptors in the intraocular pressure-lowering effects of WIN55212-2. *J Pharmacol Exp Ther* 292, 136–139.
- Stella, V. J., Rao, V. M., Zannou, E. A., & Zia, V. (1999). Mechanisms of drug release from cyclodextrin complexes. Adv Drug Delivery Rev 36, 3-16.
- Straiker, A. J., Maguire, G., Mackie, K., & Lindsey, J. (1999a). Localization of the cannabinoid CB1 receptors in human anterior eye and retina. *Invest Ophthalmol Vis Sci* 40, 2442–2448.
- Straiker, A., Stella, N., Piomelli, D., Mackie, K., Karten, H. J., & Maguire, G. (1999b). Cannabinoid CB1 receptors and ligands in vertebrate retina: localization and function of an endogenous signalling system. *Proc Natl Acad Sci USA 96*, 14565–14570.
- Sucher, N. J., Lipton, S. A., & Dreyer, E. B. (1997). Molecular basis of glutamate toxicity in retinal ganglion cells. *Vision Res* 37, 3483–3493.
- Sugiura, T., Kondo, S., Sukagawa, A., Nakane, S., Shinoda, A., Itoh, K., Yamashita, A., & Waku, K. (1995). 2-Arachidonoylglycerol: a possible

- endogenous cannabinoid receptor ligand in brain. *Biochem Biophys Res Commun* 215, 89-97.
- Sugrue, M. F. (1997). New approaches to antiglaucoma therapy. *J Med Chem 40*, 2793–2809.
- Sugrue, M. F., Funk, H. A., Leonard, Y., O'Neill-Davis, L., & Labelle, M. (1996). The ocular hypotensive effects of synthetic cannabinoids. *Invest Ophthalmol Vis Sci* 37, S831.
- Szejtli, J. (1988). Cyclodextrin Technology. . Dordrecht: Kluwer Academic Publishers.
- Szente, L., & Szejtli, J. (1999). Highly soluble cyclodextrin derivatives: chemistry, properties, and trends in development. Adv Drug Delivery Rev 36, 17–28.
- Thompson, D. O. (1997). Cyclodextrins-enabling excipients: their present and future use in pharmaceuticals. Crit Rev Ther Drug Carrier Syst 14, 1–328.
- Ueda, N., Kurahashi, Y., Yamamoto, S., & Tokunaga, T. (1995). Partial purification and characterization of the porcine brain enzyme hydrolyzing and synthesizing anandamide. J Biol Chem 40, 23823–23827.
- Urtti, A., & Salminen, L. (1985). A comparison between iris-ciliary body concentration and receptor affinity of timolol. *Acta Ophthalmol* 63, 16–18.
- Urtti, A., & Salminen, L. (1993). Minimizing systemic absorption of topically administered ophthalmic drugs. Surv Ophthalmol 37, 435–456.
- Waller, C. W., Benigni, Q. A., Harland, E. C., Bedford, J. A., Murphy, J. C., & ElSohly, M. A. (1984). Cannabinoids in glaucoma III: the effects of different cannabinoids on intraocular pressure in the monkey. In Agurell S., Dewey W. L., & Wllette R. E. (Eds.), *The Cannabinoids:* Chemical, Pharmacologic and Therapeutic Aspects (pp. 871–880). New York: Academic Press Inc.

- Wang, W., Sasaki, H., Chien, D.-S., & Lee, V. H. L. (1991). Lipophilicity influence on conjunctival drug penetration in the pigmented rabbit: a comparison with corneal penetration. Curr Eye Res 10, 571–579.
- Watsky, M. A., Jablonski, M. M., & Edelhauser, H. F. (1988). Comparison of conjunctival and corneal surface areas in rabbit and human. *Curr Eye Res* 7, 483–486.
- Woodward, D. F., Krauss, A.H.-P., Chen, J., Lai, R. K., Spada, C. S., Burk, R. M., Andrews, S. W., Shi, L., Liang, Y., Kedzie, K. M., Chen, R., Gil, D. W., Kharlamb, A., Archeanpong, A., Ling, J., Madhu, C., Ni, J., Rix, P., Usansky, J., Usansky, H., Weber, A., Welty, D., Yang, W., Tang-Liu, D.D.-S., Garst, M. E., Brar, B., Wheeler, L. A., & Kaplan, L. J. (2001). The pharmacology of bimatoprost (Lumigan ™). *Surv Ophtalmol 45*, S337−S345.
- Yazulla, S., Studholme, K. M., McIntosh, H. H., & Deutsch, D. G. (1999). Immunocytochemical localization of cannabinoid CB1 receptor and fatty acid amide hydrolase in rat retina. J Comp Neurol 415, 80–90.
- Yazulla, S., Studholme, K. M., McIntosh, H. H., & Fan, S.-F. (2000). Cannabinoid receptors on goldfish retinal bipolar cells: electron-microscope immunocytochemistry and whole-cell recordings. *Vis Neurosci* 17, 391–401.
- Yoles, E., Belkin, M., & Schwartz, M. (1996). HU-211, a nonpsychoactive cannabinoid, produces short- and long-term neuroprotection after optic nerve axotomy. *J Neurotrauma* 13, 49–57.
- Yu, S. P., Yeh, C., Strasser, U., Tian, M., & Choi, D. W. (1999). NMDA receptor-mediated K⁺ efflux and neuronal apoptosis. *Science* 284, 336-339.
- Zhang, M.-Q., & Rees, D. C. (1999). A review of recent applications of cyclodextrins for drug discovery. Expert Opin Ther Patents 9, 1697–1717.